Eosin Y Staining Solution, Alcoholic

Catalog No.: RA20093

Basic Information

Product name	Eosin Y Staining Solution, Alcoholic
Sizes	500 mL
Storage	RT
Shipping	RT
Validity	12 months

Product Introduction

Eosin, also known as eosin Y, is a synthetic dye belonging to the xanthene class. It appears as a pink or rose-colored powder. The molecular formula of alcoholic Eosin Y is $C_{20}H_8Br_4O_5$, with a molecular weight of 649.9. Eosin Y is most suitable for use with hematoxylin to visualize normal or pathological tissue structures.

EnkiLife Eosin Y Staining Solution (Alcoholic) is formulated with a proprietary preservative. It is easy to use and does not contain toxic reagents such as mercury or methanol. It is commonly used for staining tissue sections or cultured cells. After staining, the cytoplasm appears pink or red.

Experimental procedure

1. Sample Preparation

a) For paraffin-embedded sections:

Deparaffinize in xylene or xylene substitute for 5–10 min.

Replace with fresh xylene or substitute and deparaffinize for another 5–10 min.

Absolute ethanol: 5 min.

90% ethanol: 2 min.

70% ethanol: 2 min.

Distilled water: 2 min.

b) For frozen sections:

Rinse with distilled water for 2 min.

c) For cultured cells:

Fix with 4% paraformaldehyde for at least 10 min.

Rinse with distilled water for 2 min.

Replace with fresh distilled water and rinse again for 2 min.

2. Eosin Staining

Apply Eosin Y staining solution to the prepared samples for 0.5–5 min, adjusting the time based on staining results and experimental requirements.

If direct observation is intended, rinse twice with 70% ethanol.

If proceeding with dehydration, clearing, and mounting, rinse with 70% ethanol and continue with the following steps.

Note: For counterstaining after immunohistochemistry or other staining procedures, perform eosin staining after completion of the primary staining protocol.

3. Dehydration, Clearing, Mounting or Other Staining

a) Dehydration, clearing, and mounting:

95% ethanol: 2 min.

Fresh 95% ethanol: 2 min.

Absolute ethanol: 2 min.

Fresh absolute ethanol: 2 min.

Xylene or xylene substitute: 5 min.

Fresh xylene or substitute: 5 min.

Mount with neutral balsam or other appropriate mounting medium.

Observe under microscope; cytoplasm should appear pink or red.

b) For subsequent staining (e.g., immunofluorescence or Hoechst):

After eosin staining:

Rinse twice with 70% ethanol, 2 min each.

Soak in PBS, saline, TBS, or TBST for 5 min.

Proceed with immunofluorescence or other fluorescent staining protocols.

Notes

1. For dehydration, clearing, and mounting, users must prepare xylene or xylene substitute and mounting medium (e.g., neutral balsam).
2. Wear appropriate personal protective equipment (lab coat and gloves) during operation.
3. Use the reagent promptly after opening to ensure optimal performance.

This product is for research use only!